



KAMIYA BIOMEDICAL COMPANY

Mouse Anti-Sheep Red Blood Cell (SRBC) IgM ELISA

For the quantitative determination of SRBC-IgM in mouse serum and plasma

Cat. No. KT-573

For research use only, not for use in diagnostic procedures.

PRODUCT INFORMATION**Mouse Anti-Sheep Red Blood Cell (SRBC) IgM ELISA**
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The **K-ASSAY®** Mouse SRBC IgM ELISA is for the quantitative determination of SRBC IgM in mouse serum or plasma.

PRINCIPLE

The Mouse Anti-SRBC IgM ELISA is based on a solid phase enzyme-linked immunosorbent assay (ELISA). The assay uses detergent solubilized SRBC ghosts for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated anti-mouse IgM antibodies for detection. Test serum or plasma samples are diluted and incubated in the microtiter wells for 45 minutes. The microtiter wells are subsequently washed and HRP conjugate is added and incubated for 45 minutes. Anti-SRBC IgM molecules are thus sandwiched between immobilized SRBC antigens and the detection antibody conjugate. The wells are then washed to remove unbound HRP-labeled antibodies and TMB reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of stop solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of anti-SRBC IgM is proportional to the optical density of the test sample.

COMPONENTS

- Microtiter Plate: SRBC coated 96-well plate (12 strips of 8 wells)
- Enzyme Conjugate Solution: 11 mL
- Calibrator: Lyoph. x 2
- Diluent Buffer: 30 mL
- TMB Solution: 11 mL
- Stop Solution: 11 mL, 1N HCl
- Wash Buffer (20x): 50 mL

Materials or Equipment required but not provided

- Plate reader (450 nm)
- Micropipette and tips
- De-ionized water
- Graph paper (PC software is optional)
- Paper towels
- Polypropylene or glass tubes
- Vortex mixer
- Plate shaker/incubator
- Plate washer

STORAGE

Store at 4°C. Microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air. The kit is stable until the expiration date when stored as noted in this section.

General Instructions

1. Please read and understand the instructions thoroughly before using the kit.
2. This kit is designed to measure anti-SRBC IgM levels in mouse serum or plasma collected 5 days after immunization with SRBC. At this point the immune response originates almost exclusively from IgM.
3. All reagents should be allowed to reach room temperature (18-25°C) before use.
4. The optimal sample dilution should be determined empirically. However, studies suggest an initial sample dilution of 50 fold.
5. Serum and plasma samples must be diluted at least 15 fold in diluent.
6. Optimum results are achieved if, at each step, reagents are pipetted into wells of the microtiter plate within 5 minutes.

PREPARATION OF REAGENTS

Wash Buffer

The wash solution is provided as 20x stock. Prior to use dilute the contents of the bottle (50 mL) with 950 mL of distilled or deionized water.

Calibrator

1. The Mouse anti-SRBC IgM calibrator is provided as lyophilized stock. Reconstitute with volume of diluent indicated on the vial label to give you the 100 u/mL calibrator.
2. Label 5 polypropylene or glass tubes as 50, 25, 12.5, 6.25, and 3.125 u/mL, and pipette 250 μ L of diluent into each tube.
3. Into the tube labelled 50 u/mL prepare a 50 u/mL stock by mixing 250 μ L of the 100 u/mL calibrator with 250 μ L of diluent.
4. Dispense 250 μ L of diluent into the tubes labelled 50, 25, 12.5, 6.25, and 3.125 u/mL.
5. Prepare a 50 u/mL calibrator by diluting and mixing 250 μ L of the 100 u/mL calibrator with 250 μ L of diluent in the tube labelled 50 u/mL.
6. Similarly prepare the 25, 12.5, 6.25, and 3.125 u/mL calibrators by serial dilution.

SAMPLE PREPARATION

Note: Studies indicate that anti-SRBC IgM is present in mouse serum or plasma at concentrations in excess of 2,000 u/mL. In order to obtain values within range of the calibration curve, we suggest samples initially be diluted 50 fold using the following procedure for each sample tested. Optimal dilutions may need to be determined empirically.

1. For each test sample dispense 294 μ L of diluent into separate tubes.
2. Pipette and mix 6 μ L of the serum/plasma sample into the tube containing 294 μ L of diluent. This provides a 50 fold diluted sample.
3. Repeat this procedure for each sample to be tested.
4. Do not use dilutions lower than 15 fold.

PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 μ L of calibrators, and diluted samples into the wells (we recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 μ L/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 μ L of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
8. Wash as detailed in 4 and 5 above.
9. Dispense 100 μ L of TMB reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature for 20 minutes.
11. Stop the reaction by adding 100 μ L of Stop Solution to each well.
12. Gently mix. It is important to make sure all the blue color changes to yellow.
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.

CALCULATION OF RESULTS

1. Calculate the average absorbance values for each set of calibrators, and samples.
2. Construct a calibration curve by plotting the mean absorbance obtained from each calibrator against its concentration in u/mL on linear graph paper, with absorbance values on the vertical or Y axis and concentrations on the horizontal or X axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of anti-SRBC IgM in u/mL from the calibration curve.
4. Multiply the derived concentrations by the dilution factor to determine the actual concentration for anti-SRBC IgM in the serum/plasma sample.
5. PC graphing software may be used for the above steps.
6. If the OD values of samples fall outside the calibration curve when tested at a dilution of 100, samples should be diluted appropriately and re-tested. (Do not use dilutions lower than 15 fold.)

Limitations of the Procedure

- Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
- The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

FOR RESEARCH USE ONLY**KAMIYA BIOMEDICAL COMPANY**

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